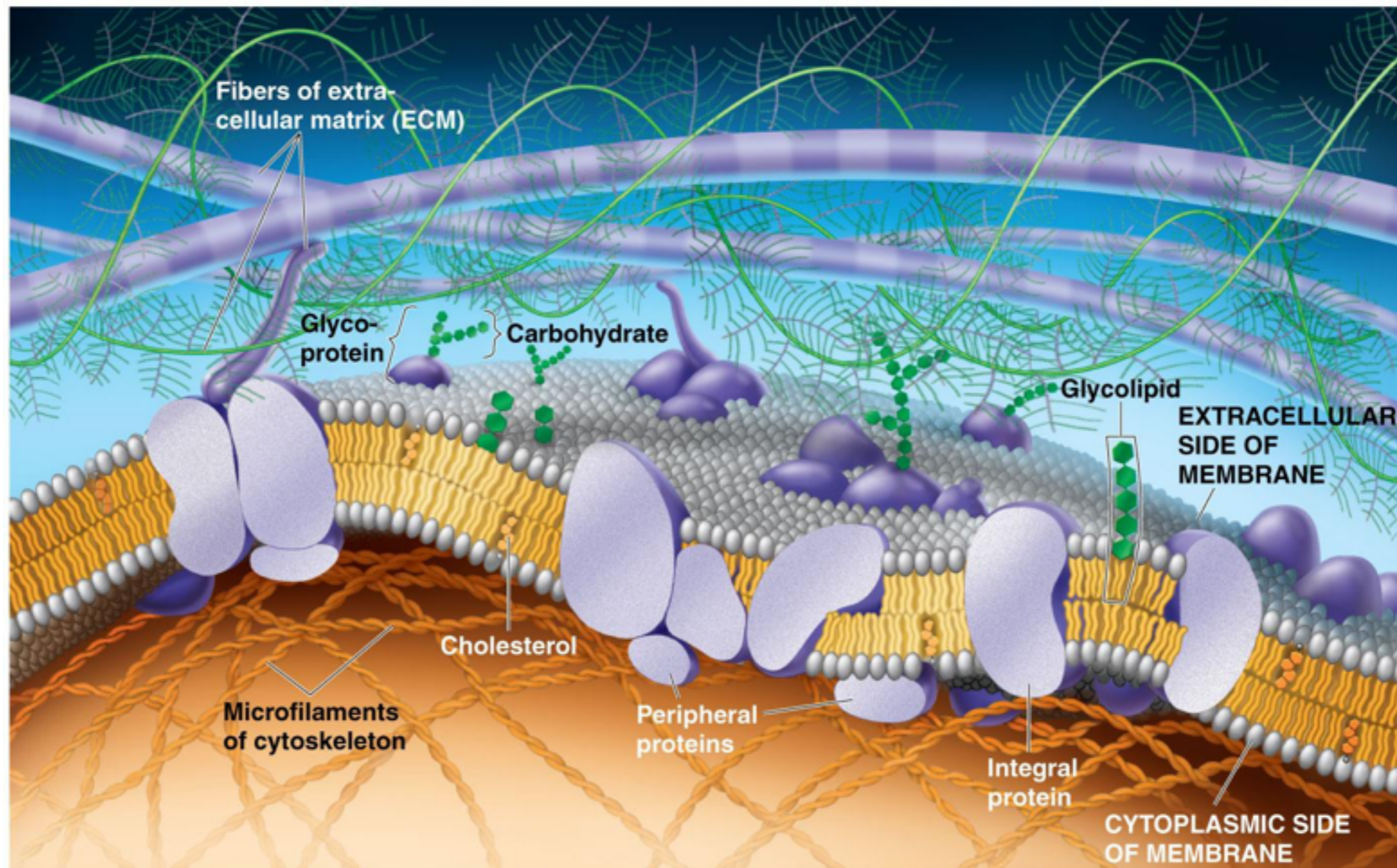
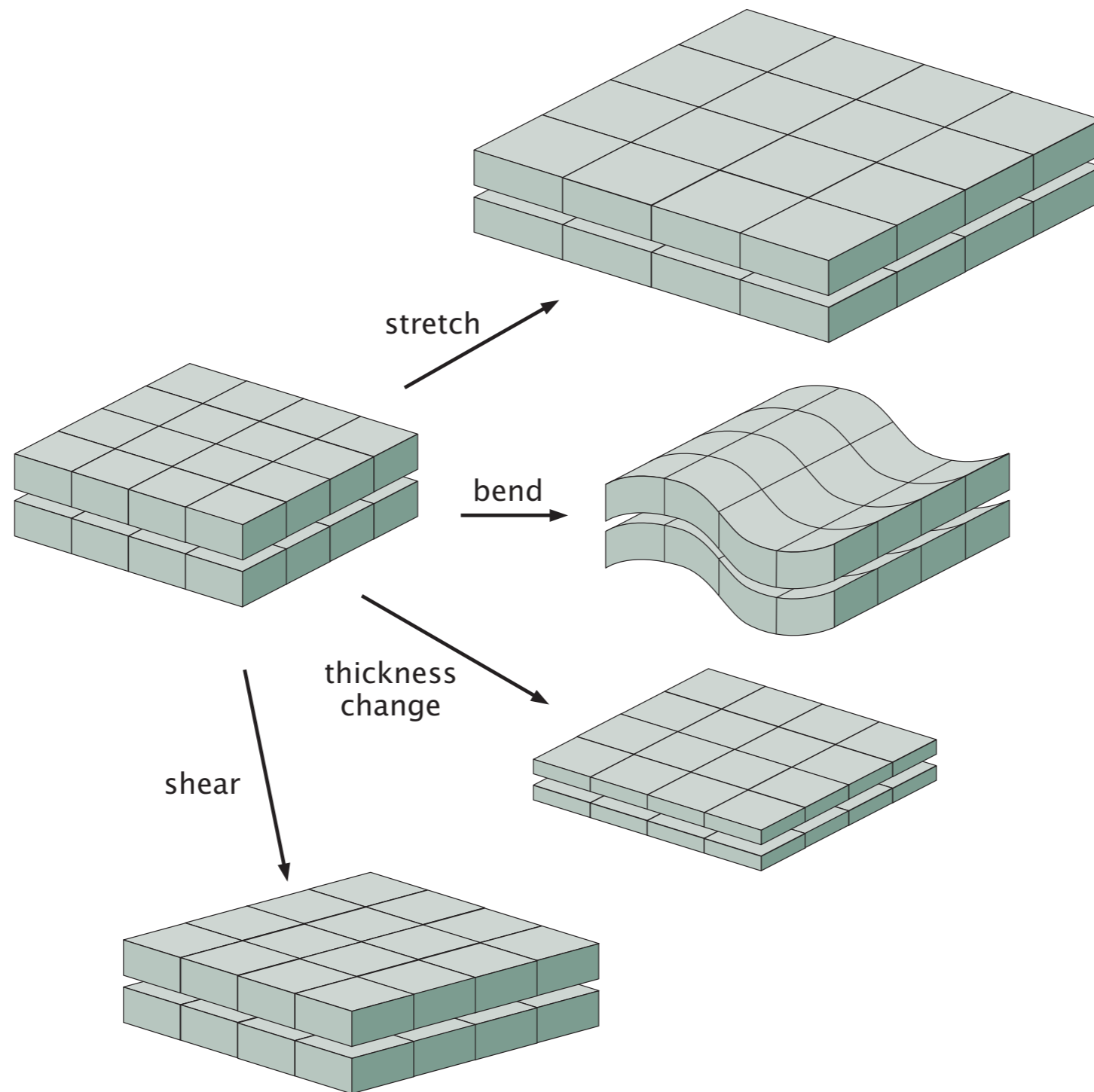


MAE 545: Lecture 16 (11/17)

Mechanics of cell membranes



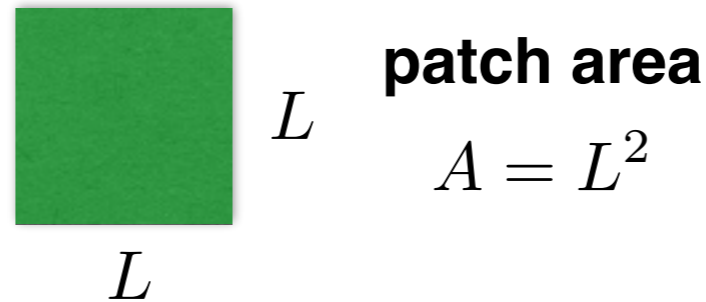
Membrane deformations



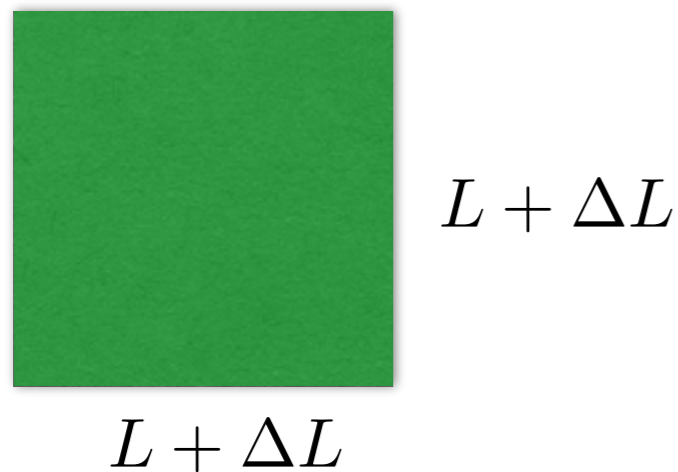
R. Phillips et al., Physical
Biology of the Cell

Energy cost for stretching and shearing

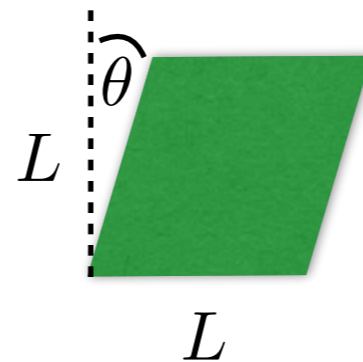
undeformed
square patch



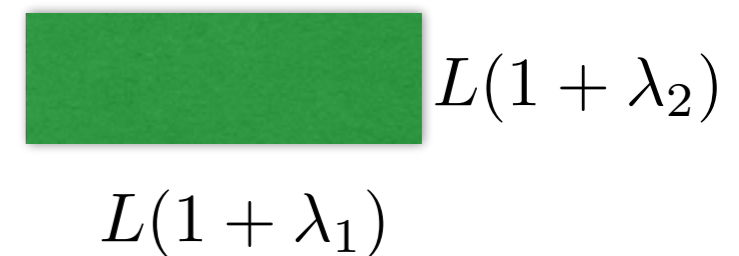
isotropic
deformation



shear
deformation



anisotropic
stretching



$$\frac{E}{A} = \frac{B}{2} \left(\frac{\Delta A}{A} \right)^2 \approx \frac{B}{2} \left(\frac{2\Delta L}{L} \right)^2$$

$$\frac{E}{A} = \frac{\mu\theta^2}{2}$$

$$\frac{E}{A} \approx \frac{B}{2} (\lambda_1 + \lambda_2)^2 + \frac{\mu}{2} (\lambda_1 - \lambda_2)^2$$

bulk modulus

$$B \sim 0.2 \text{ N/m}$$

(lipid bilayer)

shear modulus

$$\mu \sim 10^{-5} \text{ N/m}$$

(spectrin network)

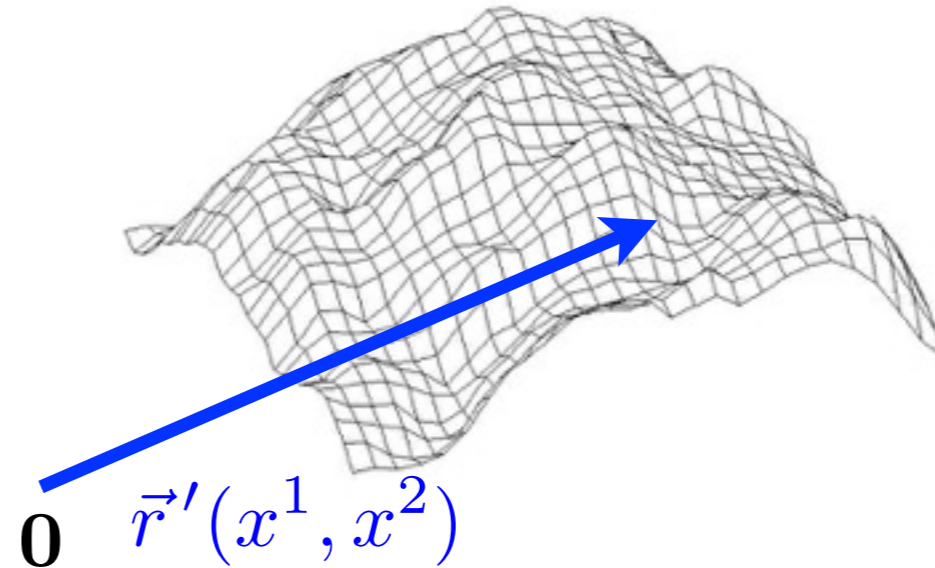
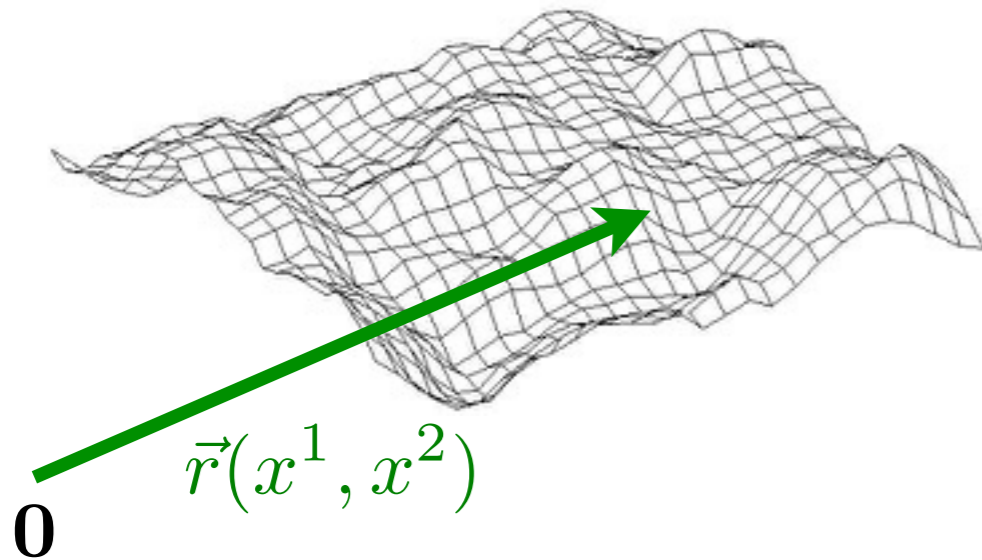
$$\lambda_1, \lambda_2 \ll 1$$

(shearing can be interpreted
as anisotropic stretching)

Strain tensor for deformation of membranes

undeformed membrane

deformed membrane



$$g_{ij} = \frac{\partial \vec{r}}{\partial x^i} \cdot \frac{\partial \vec{r}}{\partial x^j}$$

$$d\ell^2 = \sum_{i,j} g_{ij} dx^i dx^j$$

strain tensor

$$g'_{ij} = \frac{\partial \vec{r}'}{\partial x^i} \cdot \frac{\partial \vec{r}'}{\partial x^j}$$

$$d\ell'^2 = \sum_{i,j} g'_{ij} dx^i dx^j$$

Energy cost for stretching/compressing

$$u_{ij} = \frac{1}{2} \sum_k (g^{-1})_{ik} (g'_{kj} - g_{kj})$$

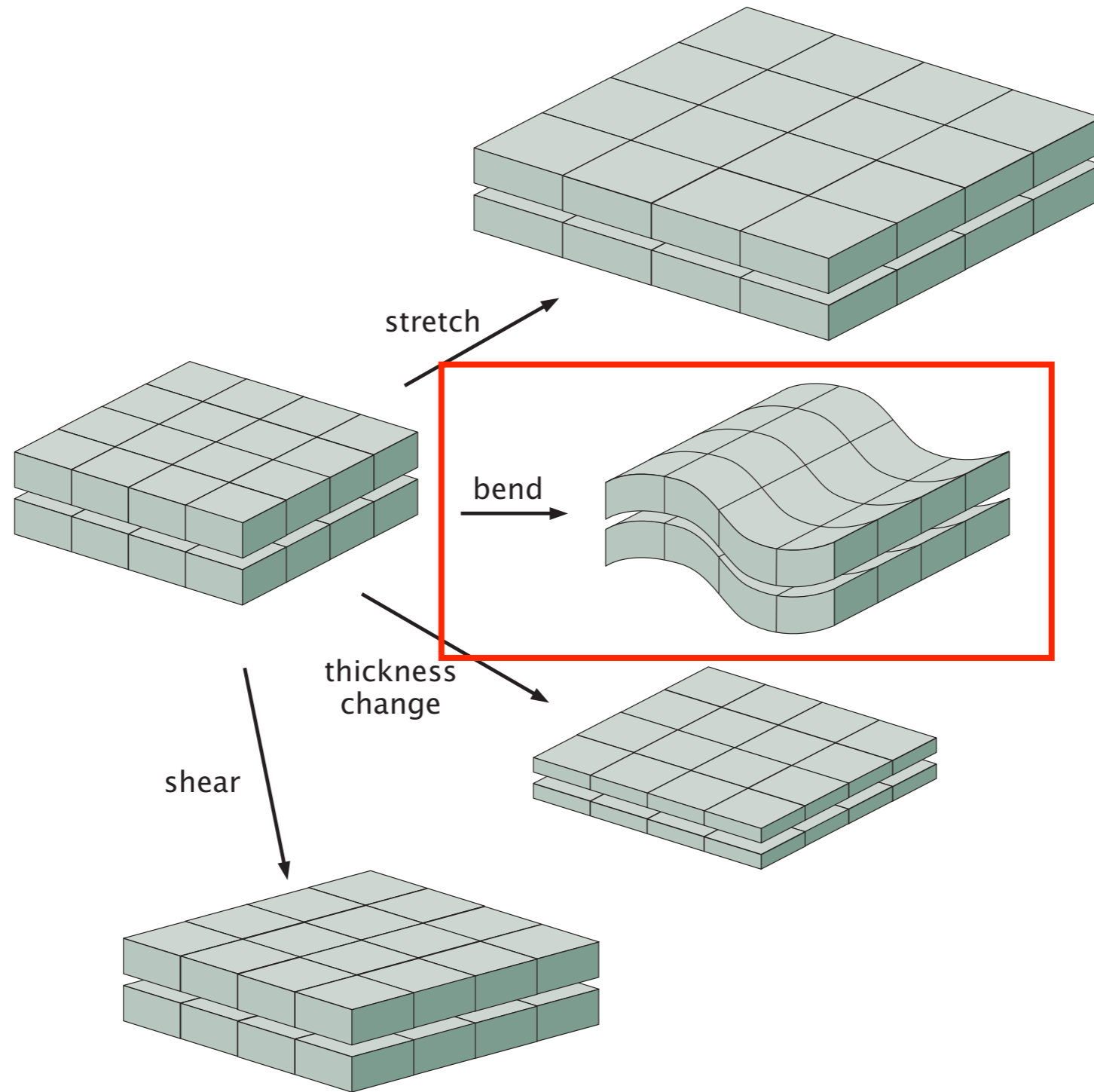
inverse metric tensor

$$\sum_k (g^{-1})_{ik} g_{kj} = \sum_k g_{ik} (g^{-1})_{kj} = \delta_{ij}$$

$$E = \int \sqrt{g} dx^1 dx^2 \frac{1}{2} \left[(B - \mu) \left(\sum_i u_{ii} \right)^2 + 2\mu \sum_{i,j} u_{ij}^2 \right]$$

$$g = \det(g_{ij})$$

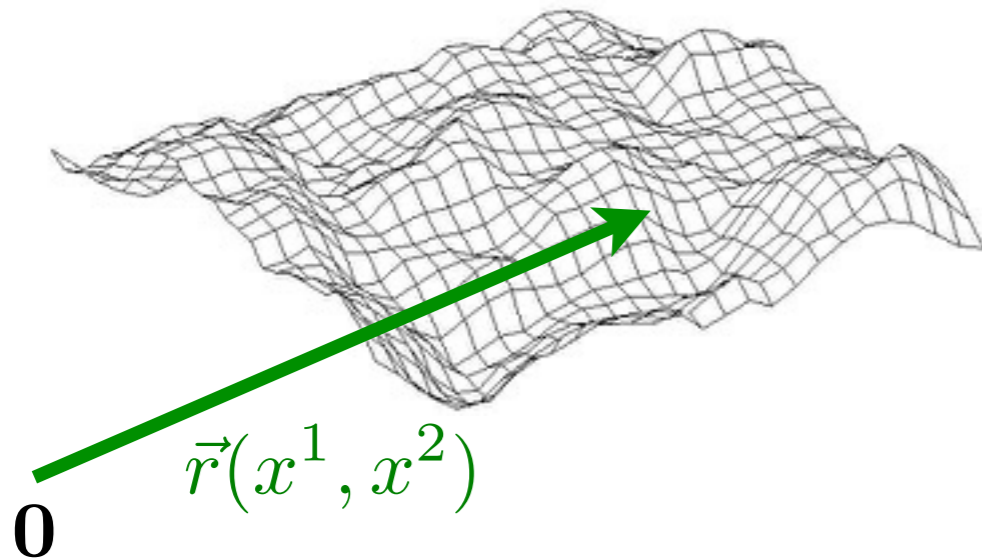
Membrane deformations



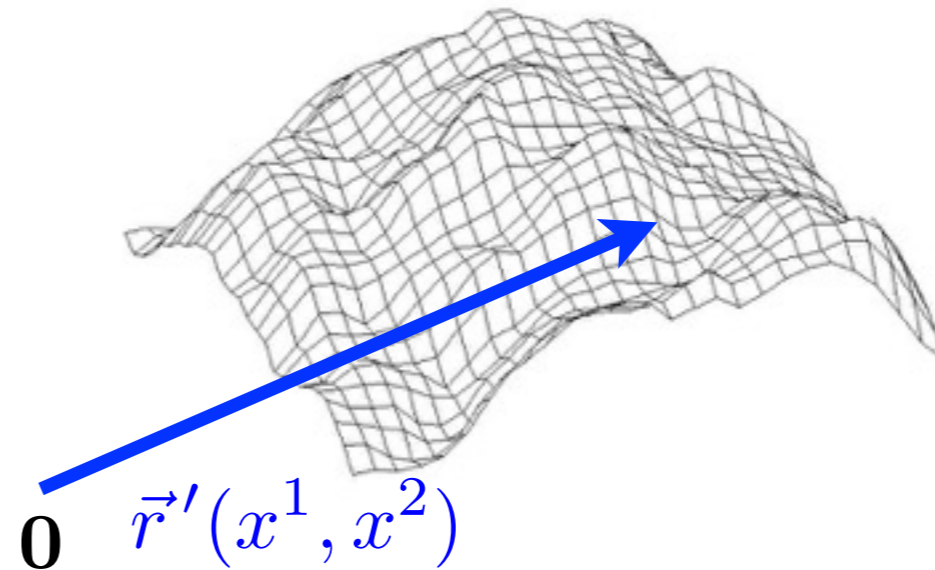
R. Phillips et al., Physical
Biology of the Cell

Bending energy for deformation of membranes

undeformed membrane



deformed membrane



$$K_{ij} = \sum_k (g^{-1})_{ik} \left(\vec{n} \cdot \frac{\partial^2 \vec{r}}{\partial x^k \partial x^j} \right)$$

$$K'_{ij} = \sum_k (g'^{-1})_{ik} \left(\vec{n}' \cdot \frac{\partial^2 \vec{r}'}{\partial x^k \partial x^j} \right)$$

bending strain tensor

$$b_{ij} = K'_{ij} - K_{ij}$$

(local measure of deviation from preferred curvature)

Energy cost of bending

$$E = \int \sqrt{g} dx^1 dx^2 \left[\frac{1}{2} \kappa \text{tr}(b_{ij})^2 + \kappa_G \det(b_{ij}) \right]$$

Bending energy

$$E = \int dA \left[\frac{\kappa}{2} \left(\frac{1}{R_1} + \frac{1}{R_2} - C_0 \right)^2 + \frac{\kappa_G}{R_1 R_2} \right]$$

**Helfrich
free energy**

bending rigidity $\kappa \sim 20k_B T$

mean curvature $H = \frac{1}{2} \left(\frac{1}{R_1} + \frac{1}{R_2} \right)$

**Gaussian
bending rigidity** $\kappa_G \sim -0.8\kappa$

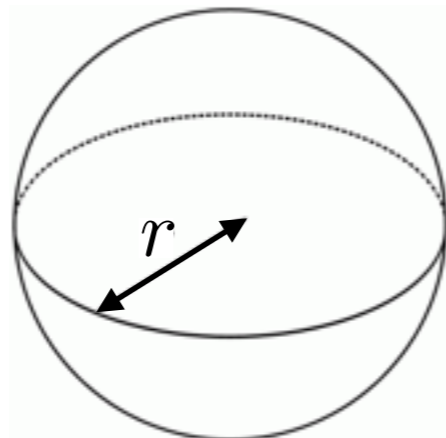
**Gaussian
curvature** $G = \frac{1}{R_1 R_2}$

**spontaneous
curvature** C_0

Example: bending energy for a sphere

$$\frac{1}{R_1} = \frac{1}{R_2} = \frac{1}{r}$$

$$C_0 = 0$$



$$E = 4\pi (2\kappa + \kappa_G) \sim 300k_B T$$

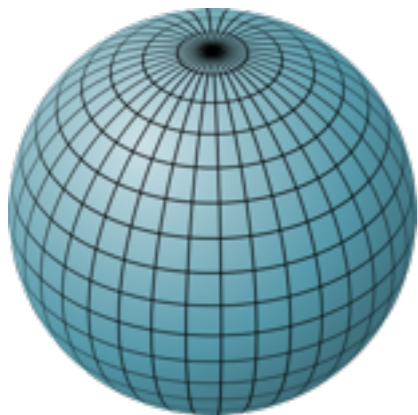
**bending energy is independent
of the sphere radius!**

Gauss-Bonnet theorem

For closed surfaces the integral over Gaussian curvature only depends on the surface topology!

$$\int \frac{dA}{R_1 R_2} = 4\pi (1 - g)$$

$g = 0$



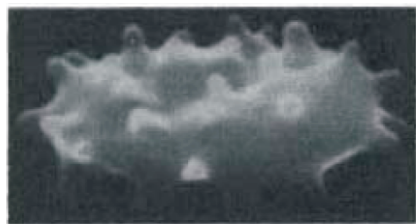
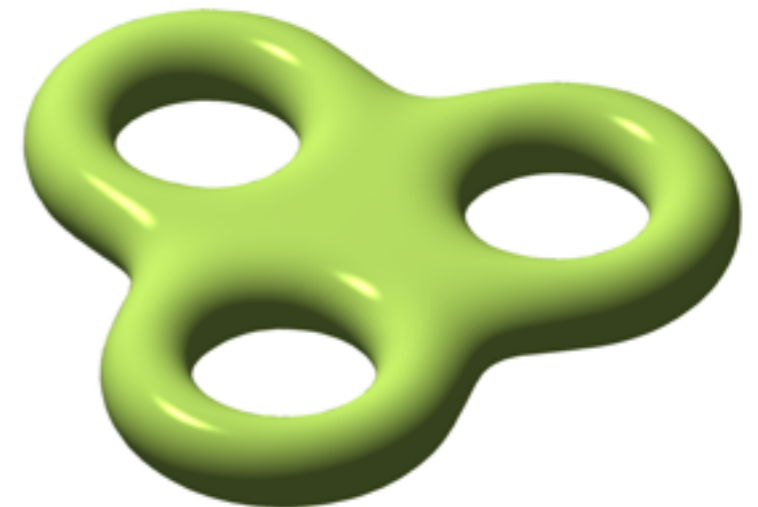
$g = 1$



$g = 2$



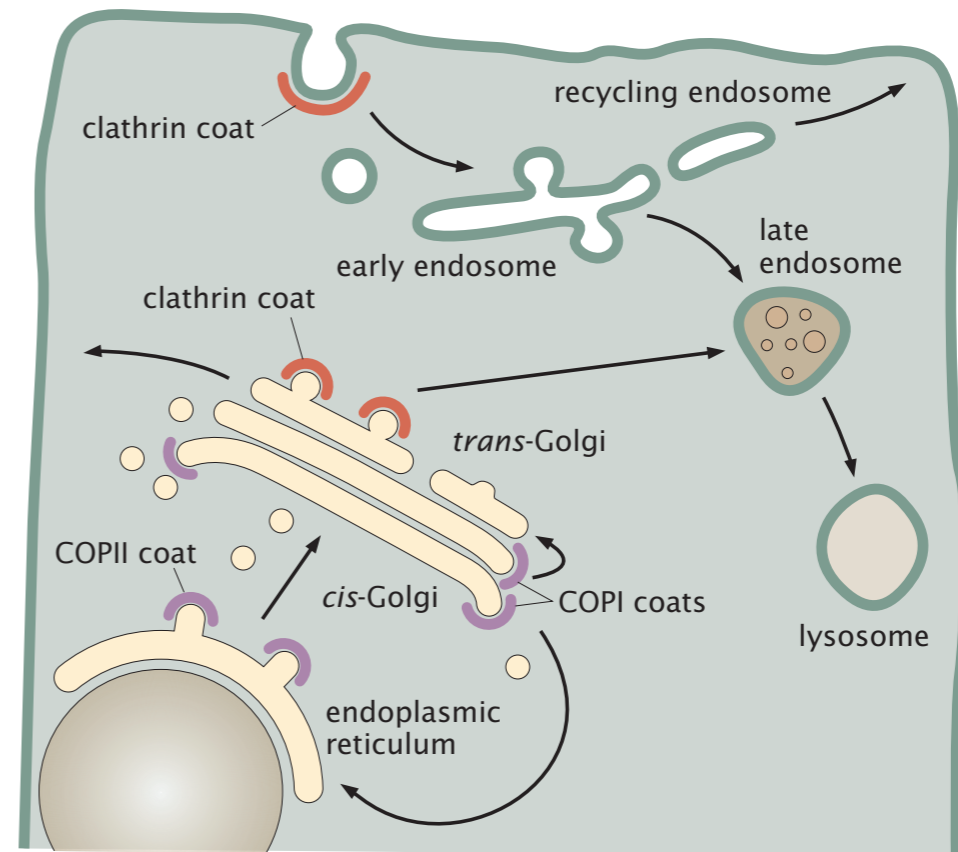
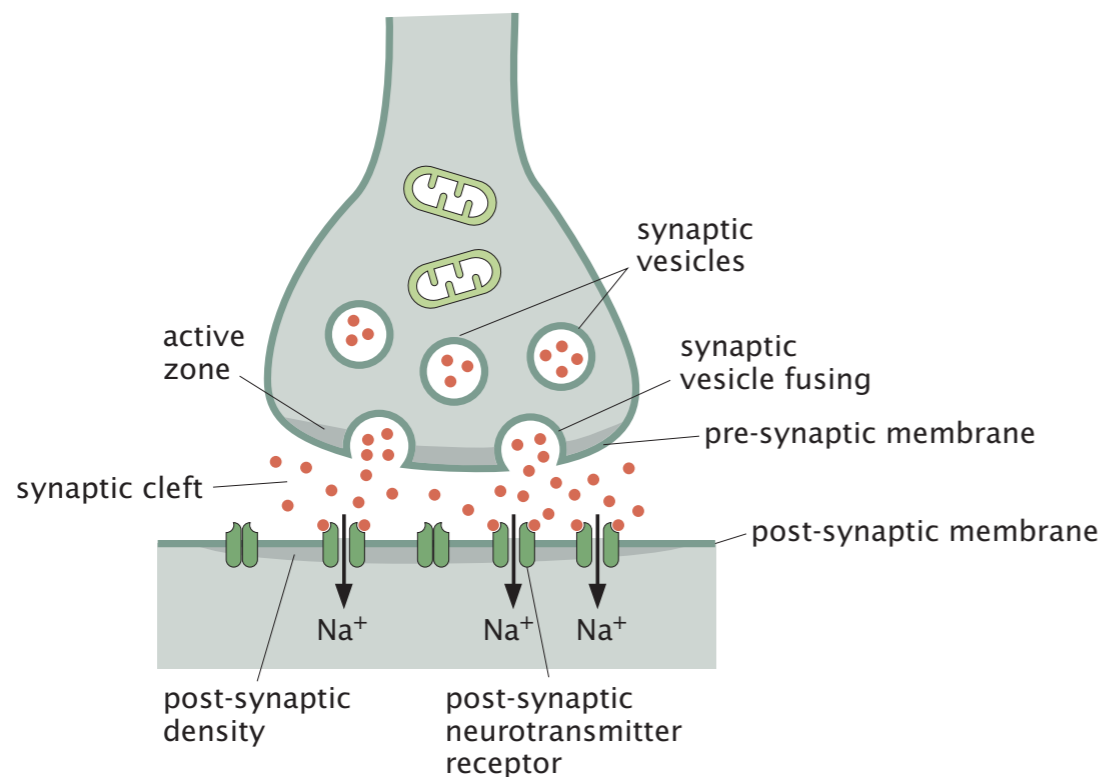
$g = 3$



It is hard to experimentally measure the Gaussian bending rigidity for cells, because cell deformations don't change the topology!

Small vesicles are used for cellular transport of molecules

transport of neurotransmitters in neuron cells

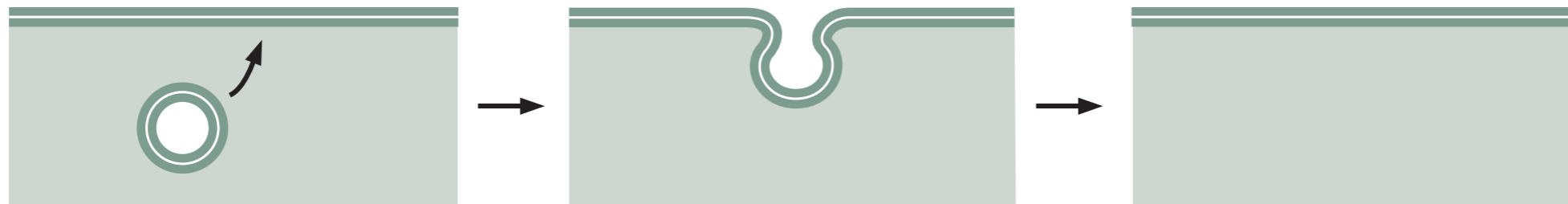


Vesicles are changing membrane topology!

R. Phillips et al., Physical Biology of the Cell

Membrane fusion

Fusion of small vesicles with the membrane is energetically favorable, but the initial merging provides a large energy barrier!



$$E = 4\pi (2\kappa + \kappa_G)$$

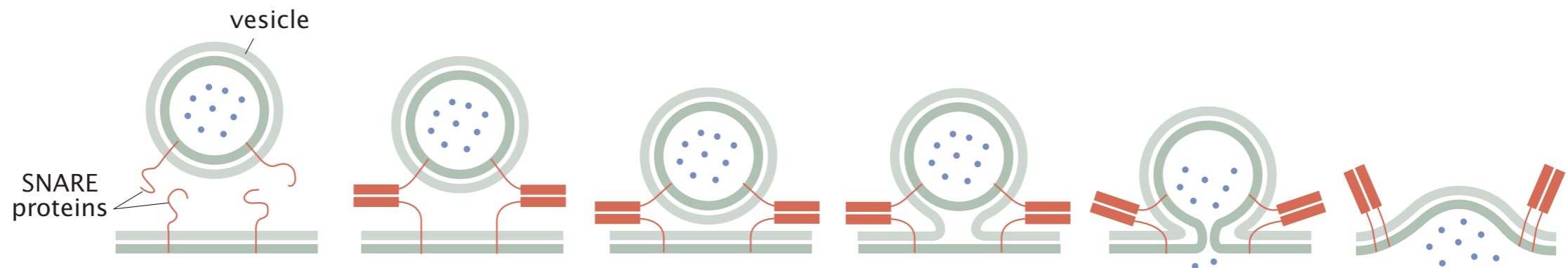
$$E \sim +300k_B T$$

$$E \approx 8\pi\kappa$$

$$E \sim +500k_B T$$

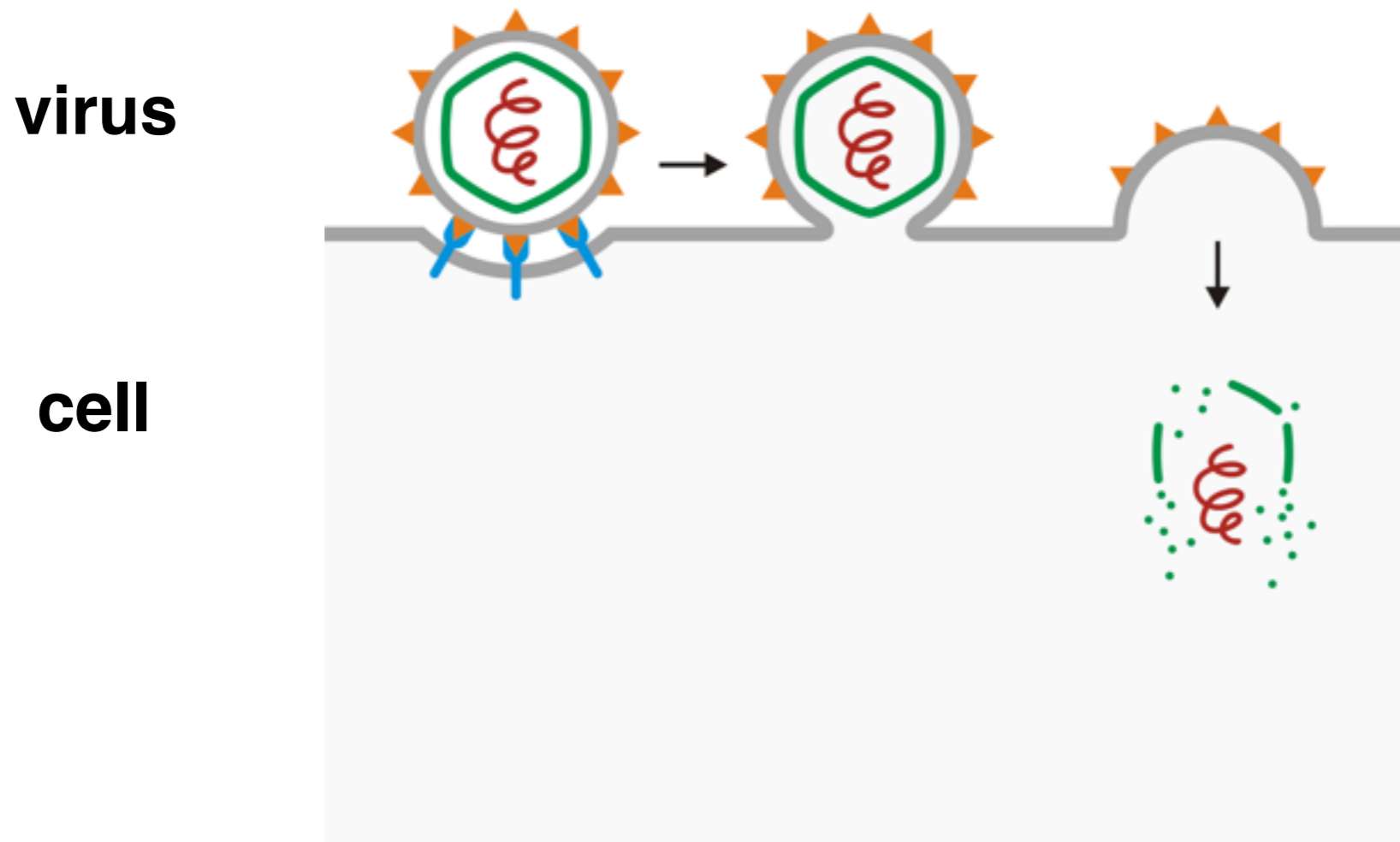
$$E = 0$$

In eukaryotic cells SNARE proteins accelerate membrane fusion by bringing vesicles closer to the membrane!



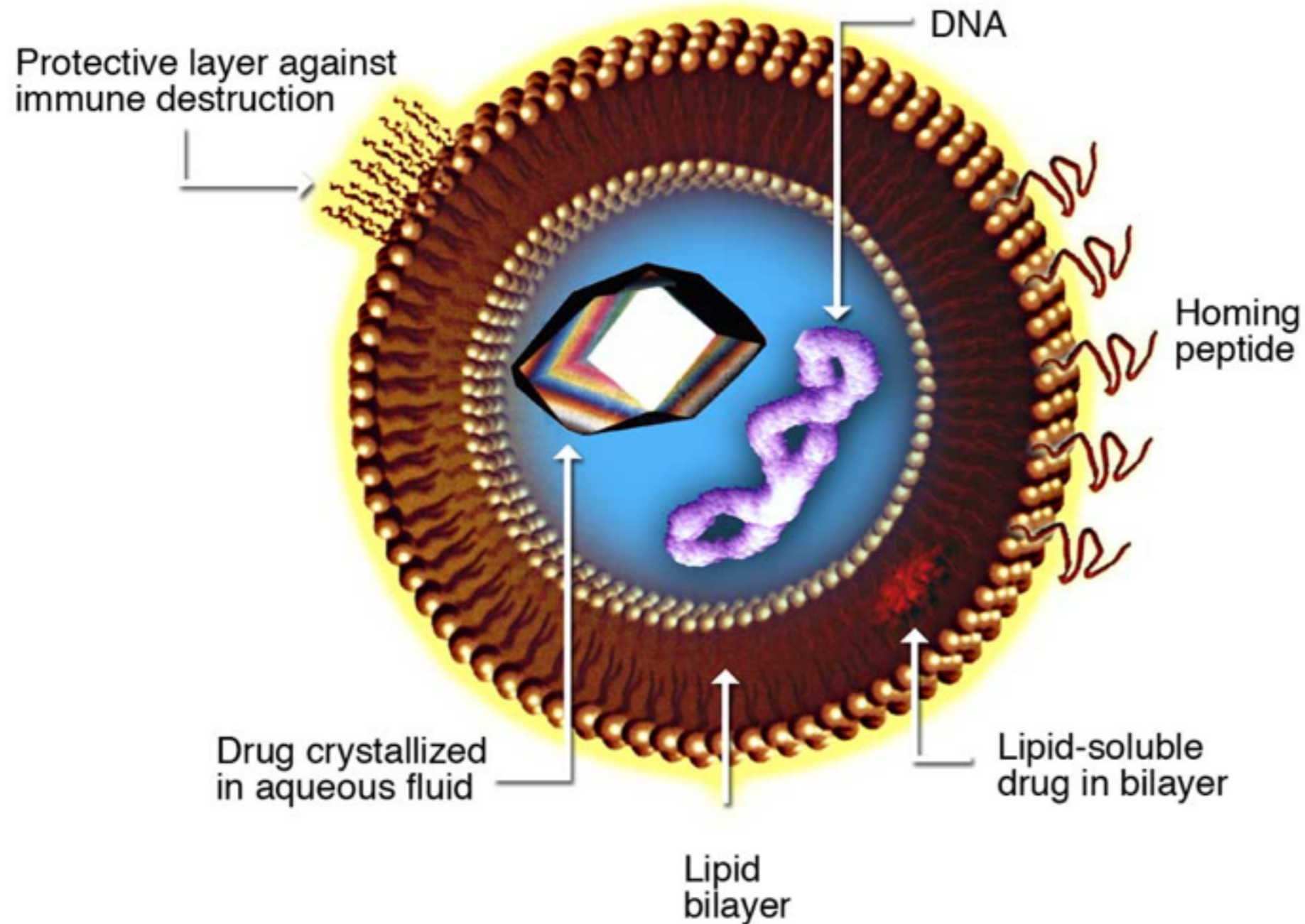
R. Phillips et al., Physical
Biology of the Cell

Viral entry to cell via receptor mediated membrane fusion



**Example of viruses with viral envelope (lipid bilayer):
HIV, influenza, hepatitis B virus, herpes viruses, ...**

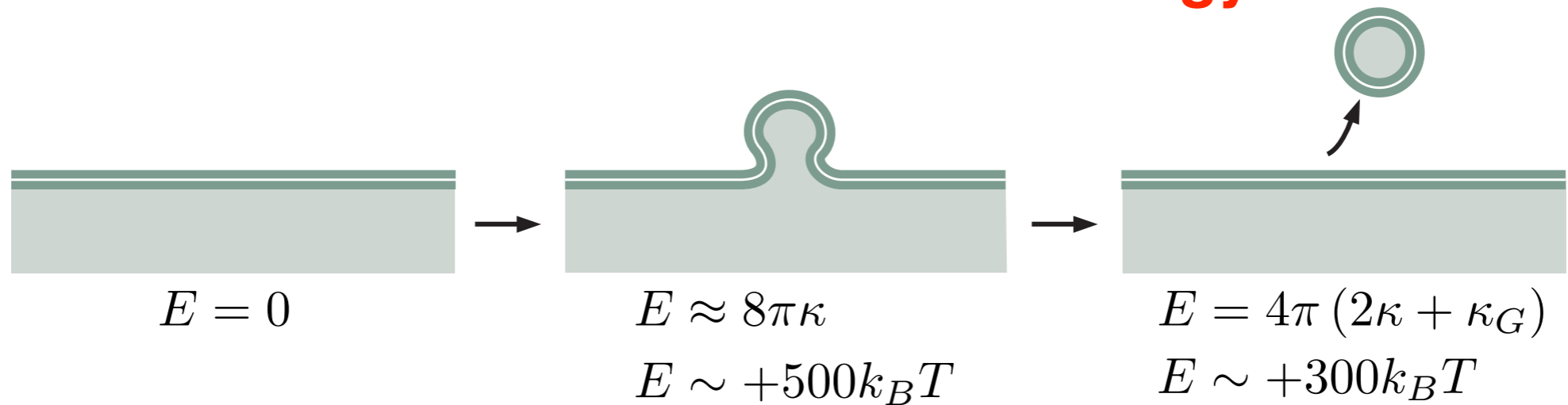
Lipid vesicles can be used for administration of drugs and nutrients



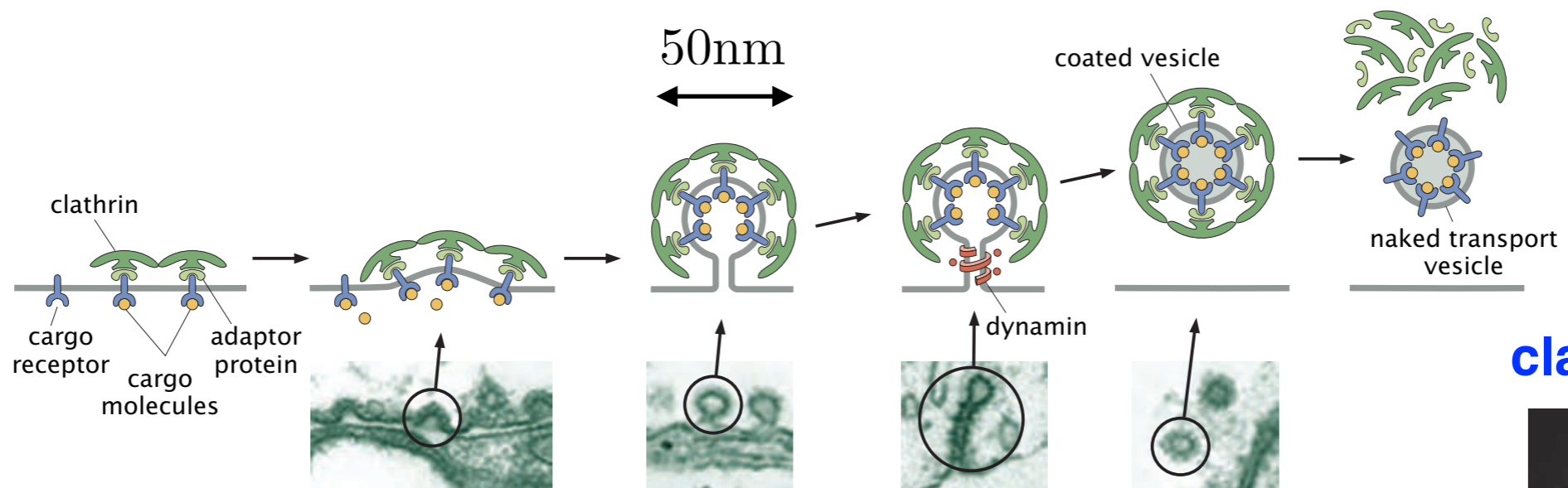
peptides bind to receptors expressed on the surface of target cells

Membrane budding

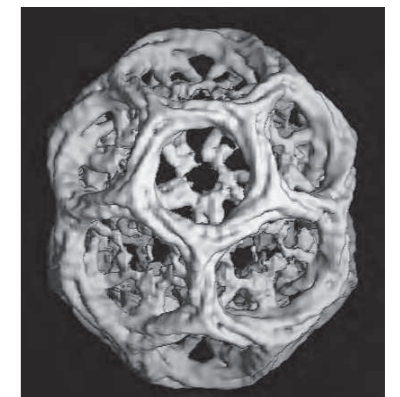
Creation of new vesicles costs energy!



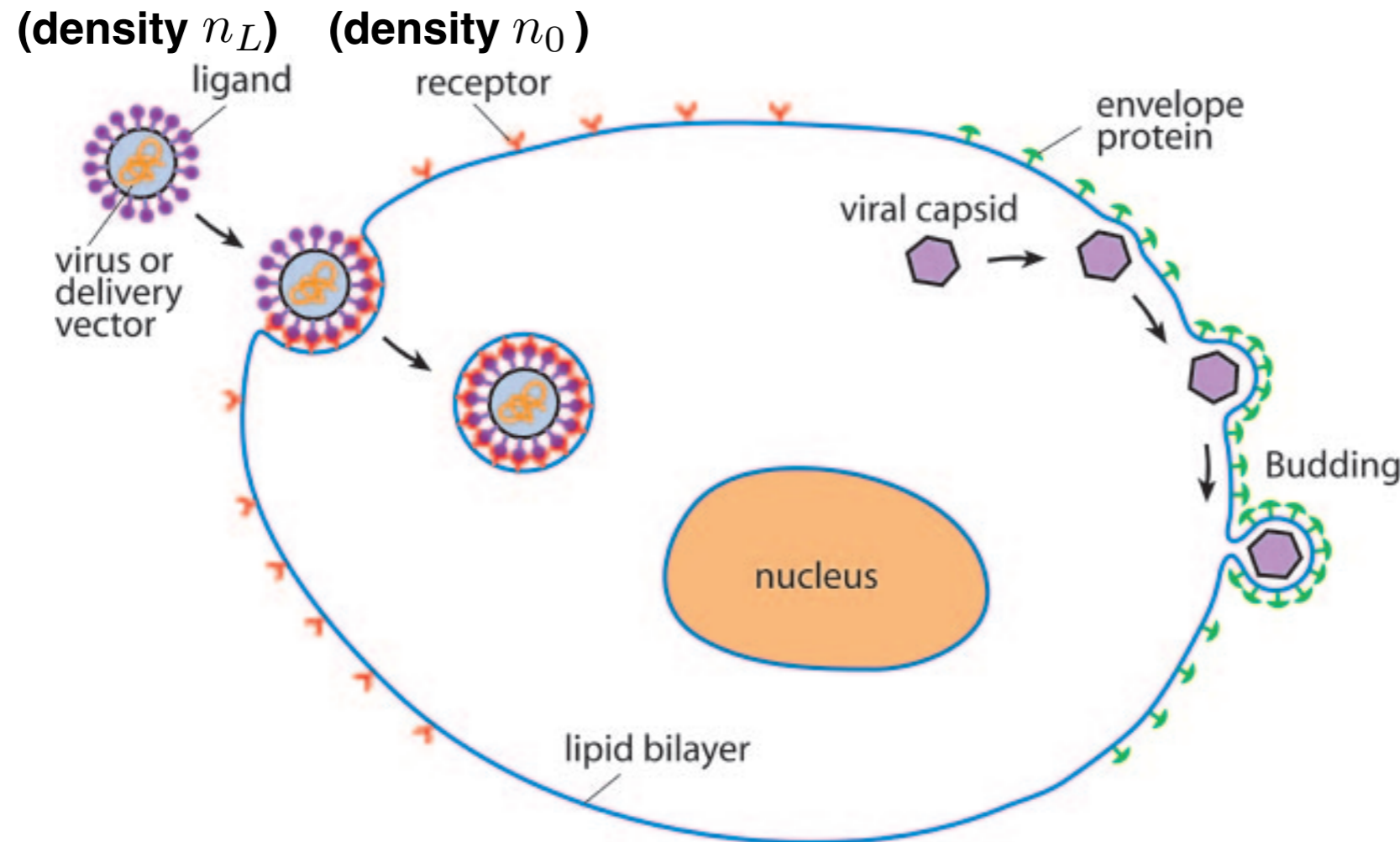
Creation of new cargo vesicles is assisted with receptor mediated coating of proteins (clathrin, COPI)



clathrin cage



Viral entry to cell via receptor mediated endocytosis



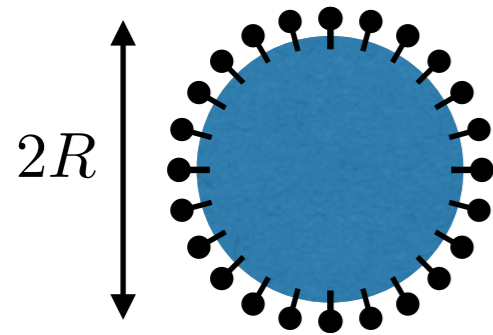
(similar process may help during budding of enveloped viruses)

Bending energy cost and loss of entropy for receptors is compensated by the binding energy between cell receptors and ligands on the surface of viral capsid.

G. Bao and X.R. Bao,
PNAS 102, 9997 (2005)

Viral entry to cell via receptor mediated endocytosis

H. Gao *et al.*, PNAS
102, 9469 (2005)



$n_L \sim 5000 \mu\text{m}^{-2}$
density of ligands



$n_0 \sim 50-500 \mu\text{m}^{-2}$
density of receptors

receptor-ligand
binding energy

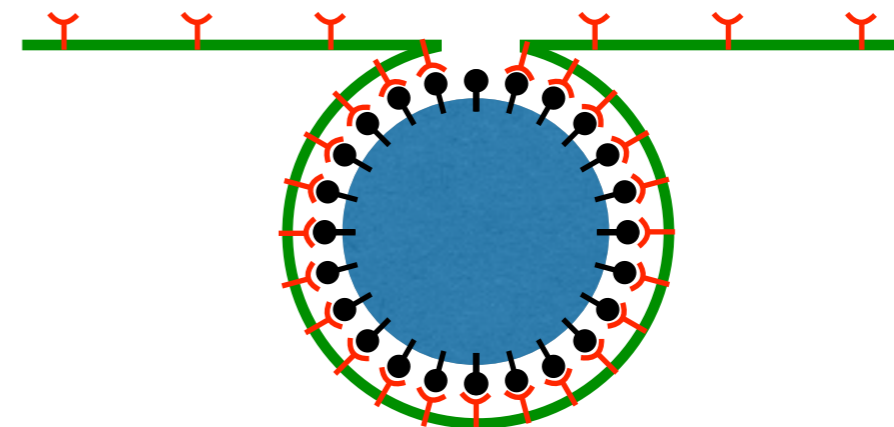
$$U_b \sim 15k_B T$$

bending rigidity

$$\kappa \sim 20k_B T$$

total number of ligands

$$N_L = 4\pi R^2 n_L$$



$$\Delta E \approx 8\pi\kappa - 4\pi R^2 n_L U_B + 4\pi R^2 k_B T n_L \ln(n_L/n_0)$$

membrane
bending
energy

binding
energy of
receptors

loss of entropy
for receptors

**Endocytosis occurs
when $\Delta E < 0$:**

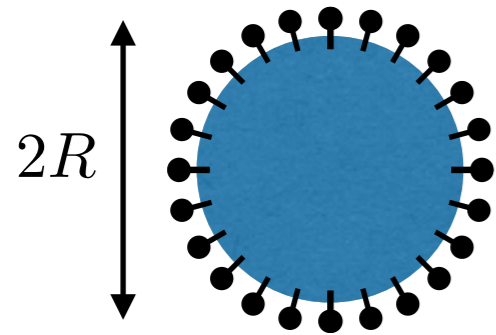


$$R > \sqrt{\frac{2\kappa}{n_L (U_B - k_B T \ln(n_L/n_0))}} \sim 30\text{nm}$$

How fast is this process?

Viral entry to cell via receptor mediated endocytosis

H. Gao *et al.*, PNAS
102, 9469 (2005)



$n_L \sim 5000 \mu\text{m}^{-2}$
density of ligands



$n_0 \sim 50-500 \mu\text{m}^{-2}$
density of receptors

receptor-ligand
binding energy

$$U_b \sim 15k_B T$$

bending rigidity

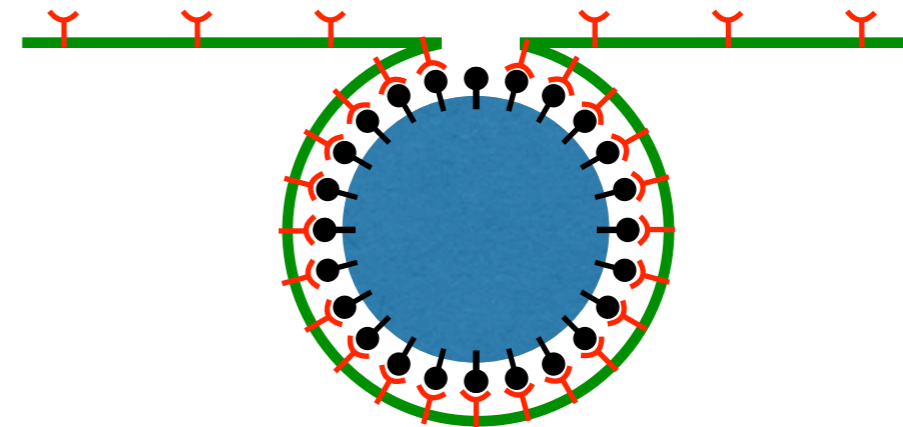
$$\kappa \sim 20k_B T$$

total number
of ligands

$$N_L = 4\pi R^2 n_L$$

diffusion of
receptors

$$D \sim 10^4 \text{nm}^2/\text{s}$$



$$R > \sqrt{\frac{2\kappa}{n_L (U_B - k_B T \ln(n_L/n_0))}} \sim 30 \text{nm}$$

**Need to recruit N_L receptors
from circular region of
radius L via diffusion**

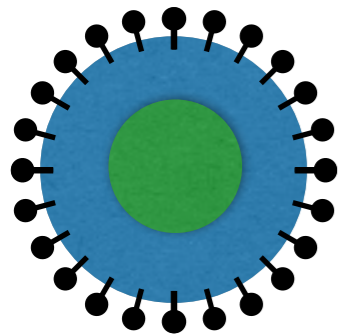
$$N_L = \pi L^2 n_0 = 4\pi R^2 n_L$$



$$t \sim \frac{L^2}{D} \sim \frac{R^2 n_L}{D n_0} \gtrsim 10 \text{s}$$

Use of magnetic nanoparticles for diagnostic and treatment of tumors

Receptors for LHRH hormone are over-expressed in breast, ovarian, and prostate cancer cells

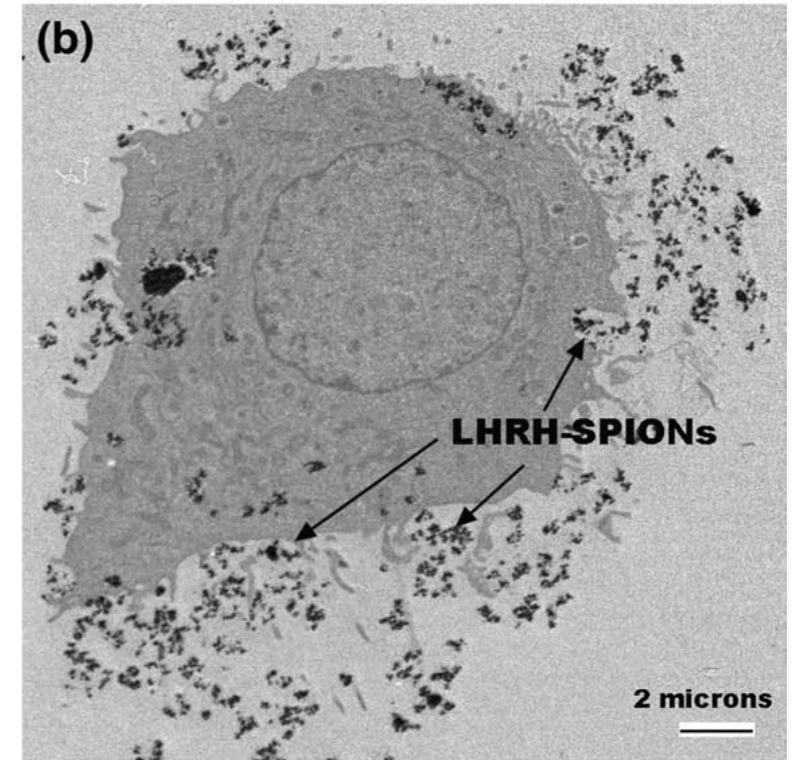


LHRH hormone
PEG coating
magnetic core

Magnetic particles enter only cancer cells via LHRH-receptor mediated endocytosis

PEG coating shields nanoparticles from immune system and prevents macro-clustering of nanoparticles.

Cancer cells containing magnetic nanoparticles can be detected with MRI (magnetic resonance imaging). Then magnetic particles can be heated via magnetic field to destroys cancer cells.



W. Soboyejo *et al.*