MAE 545: Lecture 14 (3/29) Shapes of vesicles and cells



Cells in hypotonic and hypertonic solutions

$c_{\rm in} > c_{\rm out}$ hypotonic solution

 $c_{\rm out}$

 $\frac{R}{4B}k_BT\left(c_{\rm in}-c_{\rm out}\right)$



 $R\Delta p$

4B

 $2R + 2\Delta R$

 ΔR

R

 $c_{\rm in} < c_{\rm out}$ hypertonic solution



Water flows out of the cell until concentrations become equal.

Thin cell membrane prefers to bend rather than compress

 $c_{\rm in} = c_{\rm out}$

How can we estimate the shape of "deflated" cells?

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 V_0 = $c_{\rm out}$

Area difference between lipid layers

Length difference for 2D example on the left

$$\Delta \ell = \ell_{\text{out}} - \ell_{\text{in}} = (R + w_0/2)\varphi - (R - w_0/2)\varphi$$
$$\Delta \ell = w_0\varphi = \frac{w_0\ell}{R}$$

Area difference between lipid layers in 3D

$$\Delta A = A_{\text{out}} - A_{\text{in}} = w_0 \int dA \left(\frac{1}{R_1} + \frac{1}{R_2}\right)$$

Lipids can move within a given layer, but flipping between layers is unlikely. This sets a preferred area difference ΔA_0 .

Non-local bending energy

 w_0

out

 φ

R

$$E = \frac{k_r}{2Aw_0^2} \left(\Delta A - \Delta A_0\right)^2$$

 $k_r \approx 3\kappa \approx 60k_BT$

Total elastic energy for cells (vesicles)

Shape of cells (vesicles) can be obtained by minimizing the total elastic energy

this term is constant for a given topology

$$E = \int dA \left[\frac{1}{2} (B - \mu) u_{ii}^2 + \mu u_{ij}^2 + \frac{\kappa}{2} \left(\frac{1}{R_1} + \frac{1}{R_2} - C_0 \right)^2 + \frac{\kappa_G}{R_1 R_2} \right]$$
$$+ \frac{k_r}{2A_0 w_0^2} \left(\Delta A - \Delta A_0 \right)^2 + \frac{1}{2} k_B T c_{\text{out}} V_0 \left(\frac{V - V_0}{V_0} \right)^2$$

Energetically it is very costly to change the cell volume V_0 and the membrane area A_0 (large bulk modulus B)!

Introduce dimensionless quantities that would be equal to 1 for sphere

$$\begin{array}{c} \text{definition for}\\ \text{sphere radius} \\ R_0 = \sqrt{\frac{A_0}{4\pi}} \\ a = \frac{A_0}{4\pi R_0^2} = 1 \\ \end{array} \quad v = \frac{V_0}{4\pi R_0^3/3} \\ c_0 = C_0 R_0 \\ \hline \Delta a = \frac{\Delta A}{8\pi w_0 R_0} \\ \end{array} \quad e = \frac{E}{8\pi \kappa}$$

Minimal model: minimization of bending energy for lipid vesicles

Find the shape of vesicles that minimize bending energy by constraining the volume to *v*<1.



U. Seifert et al., PRA 44, 1182 (1991)

 ${\mathcal U}$

$$e = \int \frac{da}{4} \left(\frac{1}{r_1} + \frac{1}{r_2}\right)^2$$



S. Svetina and B. Zeks, Anat. Rec. 268, 215 (2002)

Bilayer couple model of vesicles

$$e = \int \frac{da}{4} \left(\frac{1}{r_1} + \frac{1}{r_2} - c_0 \right)^2 + \frac{k_r}{\kappa} \left(\Delta a - \Delta a_0 \right)^2$$

Phase diagram of vesicle shapes that minimize the free energy for $c_0 = 0, \ k_r/\kappa \to \infty$.





Shape of red blood cells

In the usual environment red blood cells have discocyte shape. Modifying cell environment can induce different shapes.





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G. Lim et al., PNAS 99, 16766 (2002)

Sickle-cell disease (anaemia)



In low oxygen environment hemoglobin proteins inside sickle cells polymerize and form long strands.

Sickle cells are much stiffer and cannot deform in order to pass through small capillaries.

Wikipedia

Protein aggregation and diseases



(B) In concentrated solution misfolded proteins tend to form aggregates.

Cells have special proteins called chaperons, which assist proteins folding into their native state and thus prevent aggregation.

Protein aggregation is a cause of many diseases (Alzheimer's, Parkinson's, ...)

Protein aggregates are associated with diseases Parkinson's disease



α-synuclein aggregates
in dopamine producing
nerve cells





Alzheimer's disease



DNA structure





Translation of mRNA



Chaperons assist with protein folding and prevent protein aggregation

ribosome translation

isolated proteins in chaperonin chambers fold into their compact native state



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Chaperons assist with disassembly of protein aggregates



chaperons: Hsp40, Hsp70, Hsp104

Under normal cell conditions, protein aggregates are small and short lived!

S. M. Doyle et al., Nat. Rev. Mol. Cell Biol. 14, 617 (2013)